PROPERTIES: solid; d 1.85 g/ml; MP 1278°C;

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**OSHA:**  $0.002 \text{ mg/m}^3$ ; Ceiling  $0.005 \text{ mg/m}^3$ VP 0 kPa (0 mm Hg) @ 25 °C

NIOSH: 0.0005 mg/m<sup>3</sup>; Carcinogen ACGIH: 0.0002 mg/m<sup>3</sup>; Carcinogen

SYNONYMS: Beryllium metal

	SAMPLING		MEASUREMENT
SAMPLER:	Filter (0.8-µm, cellulose ester membrane, 25 or 37mm)	TECHNIQUE:	Field-portable fluorescence measurement
FLOW RATE:	1 to 4 L/min	ANALYTE:	Complex of Hydroxy benzoquinoline sulfonate (HBQS) with beryllium
VOL-MIN: -MAX:	1250 L 2000 L	SAMPLE DISS	OLUTION: 1% ammonium bifluoride
SHIPMENT:	Routine	DETECTION S	OLUTION: 63.4 μM HBQS / 2.5 mM EDTA / 50.8 mM lysine monohydrochloride (pH adjusted to 12.85 with 10 M NaOH)
SAMPLE		EXCITATION A	: 380 nm
STABILITY:	stable	Detection λ ra	<b>nge</b> ; λ <sub>max</sub> : 400 – 700 nm; 475 nm
BLANKS:	2 to 10 field blanks		

CALIBRATION: Elemental beryllium in detection solution **ACCURACY** RANGE: 0.06 to 6 µg/filter ESTIMATED LOD: 0.02 µg/filter RANGE STUDIED: 0.02 to 12.0 µg/filter **PRECISION** (r): 0.021 @  $\sim$ 0.2 µg/filter (n = 15); BIAS: none identified 0.076 @ ~1.5 µg/filter (n = 6); 0.052 @  $\sim$ 3µg/filter (n = 6) **OVERALL** PRECISION (Ŝ<sub>rT</sub>): 0.057 ACCURACY: 11.6 (6.4 - 16.8, 95% C.I.) APPLICABILITY: The working range of the method is 0.06 to 6 μg/m³ for a 1,000-L air sample,. The analysis is for total beryllium

and is not compound specific.

INTERFERENCES: Minor interference (<10%) from Fe can result if iron concentrations are high. Samples high in iron demonstrate a yellow or gold coloration. This interference can be minimized by allowing the solution to sit for at least four hours, during which time the solution clears and then filtering the sample extract before use,

OTHER METHODS: Method 7300 (hot plate or microwave digestion and inductively coupled plasma atomic emission spectrometry) is an alternative (reference) procedure for the determination of elemental beryllium. ASTM method D7202-05 is a similar procedure to detect elemental beryllium by fluorescence.

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## **REAGENTS:**

- Dissolution solution: 1% Ammonium bifluoride (prepared from dissolving 1g ammonium bifluoride in 100ml of deionized water).
- Detection solution: 63.4 μM hydroxy
  benzoquinoline sulfonate (HBQS) [1] / 2.5 mM
  ethylene diamine tetraacetic acid
  disodiumdihydrate (EDTA) / 50.8 mM lysine
  monohydrochloride (pH adjusted to 12.85 with
  10 M NaOH)
- 3. Water, deionized
- Beryllium spectroscopic standard solution,
   ~1,000 µg/ml (commercially available).
- Spiked beryllium media (commercially available)

\* See SPECIAL PRECAUTIONS

## **EQUIPMENT:**

- Sampler: Membrane filters, mixed cellulose ester (MCE) or nylon, 0.8-µm pore size, 25- or 37-mm diameter
- Personal sampling pump, 1 to 4 l/min, with clamps and flexible connecting tubing
- 3. Portable ultraviolet/visible (UV/Vis) fluorometer, with irradiance excitation lamp ( $\lambda$  = 380 nm) and time-integrating visible detector (400 700 nm,  $\lambda_{max} \approx$  475 nm) or optical filters for appropriate wavelengths
- 4. Mechanical agitator, shaker or rotator
- Fluorescence cuvettes, disposable, 10-mm diameter, transparent to UV/Vis radiation
- 6. Centrifuge tubes, plastic, 15-ml
- Syringe filters, 0.45-µm nylon, 13- or 25-mm diameter, in plastic housings
   NOTE: Polytetrafluoroethylene (PTFE) filters are unsuitable for this method.
- Pipettors, mechanical, of assorted sizes as needed
- Pipet tips, plastic, disposable, of assorted sizes as needed
- Labware, plastic (e.g., beakers, flasks, graduated cylinders, etc.), of assorted sizes as needed
- 11. Tweezers, plastic or plastic-coated

12. Laboratory wipes

 Personal protective wear, e.g., respirators, masks, gloves, lab coats, safety eyewear, etc. as needed

special precautions: Wear appropriate personal protection during sampling activities and analysis. Perform sample preparation and analysis in a clean well ventilated area that is well removed from any possible beryllium contamination. It is ESSENTIAL that suitable personal protective equipment, including suitable gloves, eye protection, laboratory coat, etc. is used when working with the chemicals. Any area affected by the dissolution or the detection solution must be immediately washed with plenty of water. Ammonium bifluoride will etch glass, so it is essential that all NH<sub>4</sub>HF<sub>2</sub> solutions are contained in plastic labware. Avoid exposure by contact with the skin or eyes, or by inhalation of the vapor.

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SAMPLING:

1. Calibrate each personal sampling pump with a representative sampler in line.

- 2. Sample at an accurately known flow rate between 1 and 4 L/min for a total sample size of 500 to 2000 L for TWA measurements. Do not exceed a filter loading of ~2 mg total dust.
- After sampling, remove the filters from the cassettes using clean tweezers, and place into labeled
   15-ml plastic centrifuge tubes.

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## **SAMPLE PREPARATION:**

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4. Add 5 ml of the dissolution solution (1% ammonium bifluoride) to each 15-ml centrifuge tube containing an air filter sample, and cap each tube.

26	5.	Place each tube into a mechanical rotator, and rotate for at least 30 min.
27		NOTE: Rotator may also be substituted by a shaker or an agitator as long as the dissolution
28		solution wets the filter well. Sonication has also been shown to be effective. Dissolution of
29		refractory material such as beryllium oxide by heating the solution to 80°C for 30 minutes
30		without agitation has been shown to be effective.
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32	6.	Filter each solution with a nylon syringe filter into a clean beaker.
33	7.	Pipet 0.1 ml of each sample filtrate Into cuvettes containing 1.9 ml of the detection solution. Cap
34		and mix briefly.
35		NOTE: If high iron or titanium concentration is suspected or is evident (owing to the appearance
36		of suspended precipitate), allow the solution to settle, or filter the solution using a nylon
37		syringe filter.
38		NOTE: The stability of the detection and the dissolution solution is more than one year and of the
39		mixed measurement solution comprising both is greater than 30 days. The solutions must
40		be kept in sealed containers and the detection and mixed solutions must be stored away
41		from light.
42		NOTE: Alternative ratios of dissolution solution comprising beryllium and detection solution may
43		be used for analyzing alternative range of beryllium concentration.
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46	CALIB	RATION AND QUALITY CONTROL:
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48	8.	Calibrate the fluorometer according to the manufacturer's recommendations
49		NOTE: Beryllium stock standard solutions are made up using Beryllium spectrometric standards
50		diluted into 1% ammonium bifluoride. Calibration check standards are then prepared by
51		adding 0.1 ml of beryllium stock standards into 1.9 ml of detection solution (20-fold
52		dilution). A recommended series of standard stock solutions are 800 ppb, 200 ppb, 40
53		ppb, 10 ppb and 0 ppb to measure a range of 0.2 $\mu g$ to 4 $\mu g$ of beryllium on the sampling
54		media.

NOTE: If alternative ratios of dissolution solution comprising beryllium and detection solution are NIOSH Manual of Analytical Methods (NMAM), Fourth Edition

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56 used for sample preparation, then a similar ratio for calibration is required. 57 9. Analyze a calibration standard, a reagent blank, and a media blank at least once every 20 58 samples. Ensure that the concentration range of the calibration standards spans the measured 59 beryllium levels in the samples. Prepare a calibration graph. 60 10. Check recoveries with at least two media spikes per ten samples. 61 NOTE: If it is suspected that beryllium oxide may be present, then it is recommended to use 62 beryllium oxide for media spikes. 63 64 65 **MEASUREMENT:** 66 67 11. For each sample, obtain the fluorescence spectrum using a 380-nm excitation lamp and visible 68 (400-700 nm) detector (follow instrument manufacturer's instructions). 69 12. If the fluorescence values for any of the samples are above the range of the calibration 70 standards, dilute the sample filterate with dissolution solution, reanalyze, and apply the 71 appropriate dilution factor in subsequent calculations. 72 73 74 **CALCULATIONS:** 75 76 13. Obtain the solution concentration for each sample, C<sub>s</sub> (μg/l), and the average media blank, C<sub>b</sub> 77  $(\mu g/I)$ . 78 14. Using the solution volumes of sample,  $V_s$  (ml), and media blank,  $V_b$  (ml), calculate the concentration, C (µg/m³) of Be in the air volume sampled, V (I), while accounting for the dilution 79 80 factor DF: 81 82 83  $C = DF \times \frac{\left[C_s \ V_s - C_b \ V_b\right]}{V}, \mu g/m^3$ 84

NOTE: The table below can be used for correlating the amount of beryllium in the solution to the beryllium in the sampling media.

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Preparation of Standard	Final concentration of beryllium	Corresponding amount of
Solutions	(ppb) in calibration standard	beryllium in the media*
	solutions	
0.1 ml of <b>0 ppb standard</b> +	0.0	Corresponds to 0.00 μg Be on
1.9 ml of detection solution		media
0.1 ml of <b>10 ppb standard</b> +	0.5	Corresponds to 0.05 μg Be on
1.9 ml of detection solution		media
0.1 ml of <b>40 ppb standard</b> +	2.0	Corresponds to 0.2 μg Be on
1.9 ml of detection solution		media
0.1 ml of 200 ppb standard +	10.0	Corresponds to 1 µg Be on
1.9 ml of detection solution		media
0.1 ml of 800 ppb standard +	40.0	Corresponds to 4 µg Be on
1.9 ml of detection solution		media

<sup>\*</sup>Incorporating sample dilution factor for 5 ml of dissolution solution; note that volumes other than 5 ml will require a different appropriate dilution factor.

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## **EVALUATION OF METHOD:**

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The method was evaluated in accordance with published guidelines [3]. Experiments were conducted using an Ocean Optics® portable fluorescence device with the following components:

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- USB 200 spectrometer with spectral grating #2 (UV/Vis 600)
- 98 LS-1 lamp (380-nm) in LS-450 housing
- 99 UV-2 casting
- 100 OFLV linear filter 200-850

101	L2 collection lens and slit-200
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103	Tests were carried out in relative irradiance mode using 2- or 5-sec integration times [4].
104	
105	The method was evaluated using beryllium oxide spiked onto mixed cellulose ester (MCE) filters at levels
106	of 0, 0.02, 0.1, 0.2, 0.3, 0.4, 1.5, 3.0, and 6.0 $\mu g$ (five samples at each level).
107	
108	Long-term stability of samples was verified from spikes (n = 30) of 0.1 $\mu$ g Be on MCE filters. Samples
109	were analyzed at day one (n = 12) and then one week (n = 6), ten days (n = 3), two weeks (n = 3), three
110	weeks (n = 3), and one month (n = 3) after spiking. No diminution of fluorescence signal was observed
111	from samples prepared and analyzed after having been stored for up to thirty days.
112	
113	Interference tests were carried out using solutions of 0 nM, 100 nM, and 1.0 $\mu$ M Be in the presence of 0.4
114	mM Al, Ca, Co, Cu, Fe, Ti, Li, Ni, Pb, Sn, U, V, W, and Zn (separate experiments were carried out for each
115	potential interferant). An interlaboratory evaluation of the method was also performed [5].
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L37	
L38	
L39	METHOD DEVELOPED BY:
L40	
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L48	

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